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54 **Albumin-based nucleotides, their replication and use, and plasmids for use therein.**

57 The DNA sequence coding for human serum albumin has been isolated and inserted as two fragments into two novel plasmids which can be replicated in *E. coli*. These novel fragments can be joined to provide a unitary DNA sequence which then can be cloned into a suitable host, e.g. *E. coli*, for the expression of human serum albumin (which is used extensively in medical practice in treating shock conditions).

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ALBUMIN-BASED NUCLEOTIDES, THEIR REPLICATION
AND USE, AND PLASMIDS FOR USE THEREIN

This invention relates to nucleotides related to human serum albumin (HSA), their replication and use, and plasmids (and host substances) for use therein.

The gene for serum albumin is regulated in development. On the other hand, serum albumin is synthesised in mammals by the adult liver, and its plateau in adulthood. The embryonic liver and yolk sac, on the other hand, produce predominantly α -fetoprotein, but the synthesis decreases drastically after birth. Recently, Law et al determined the complete sequence of mouse α -fetoprotein mRNA, Nature 291 (1981) 201-205. The structure revealed extensive homology to mammalian serum albumin, indicating that the two proteins are encoded in the same gene family. Similar conclusions have been reached from studies on the α -fetoprotein genes of the rat and the mouse; see Jagodzinski et al, Proc. Natl. Acad. Sci. USA, 78 (1981) 3521-3525, and Gorin et al, J. Biol. Chem. 256 (1981) 1954-1959.

The complete nucleotide sequence of human serum mRNA has been determined from recombinant cDNA clones and from a primer-extended cDNA synthesis on the mRNA template. The sequence comprises 2,078 nucleotides, starting upstream of a potential ribosome binding site in the 5'-untranslated region. It contains all the translated codons and extends into the poly(A) at the 3'-terminus. Part of the translated sequence codes for a hydrophobic prepeptide met-lys-trp-val-thr-phe-ile-ser-leu-leu-phe-leu-phe-ser-ser-ala-tyr-ser, followed by a basic propeptide arg-gly-val-phe-arg-arg. These signal peptides are absent from mature serum albumin and, so far, have not been identified in their nascent state in humans. A remaining 1,755 nucleotides of the translated mRNA sequence code for 585 amino acids which are in agreement, with few exceptions, with the published amino acid data for human serum albumin. The mRNA sequence verifies and refines the repeating homology in the triple-domain structure of the serum albumin molecule.

DETAILED DESCRIPTION OF THE INVENTION

Human serum albumin cDNA is cloned into the PstI site of plasmid pBR322 by the oligo(dG)-oligo(dC) tailing technique. Plasmid DNA was isolated from 97 positive colonies which hybridized to the enriched albumin cDNA probe, and the recombinant plasmid pHA36 was found to contain the largest insert of an albumin cDNA sequence. Its restriction endonuclease map is shown in the drawing, together with a restriction map of the primer-extended plasmid clone pHA206. The latter was obtained in a second transformation experiment after initiating the cDNA synthesis from an internal primer. This primer was a 91 base pairs long DNA fragment, MspI(152)-TaqI(182/3), isolated from pHA36. The two plasmids, pHA36 and pHA206, share 0.15 kb of homologous DNA. Together, they encode the entire sequence for human serum albumin, starting with the CTT codon for leu -10 of the prepeptide and extending into the 3'-untranslated region of poly(A).

Sequence of the Albumin cDNA. The sequence was determined for the most part on both DNA strands to ensure accuracy. All of the restriction sites used to end-label DNA fragments were sequenced across by labeling a neighboring restriction site. The entire nucleotide sequence of the serum albumin mRNA, as determined from the cloned DNA in pHA36, pHA206, and from the primer-extended cDNA at the 5'-terminus of the message, is shown in the following Table 1. The inferred amino acid sequence is also indicated. The mRNA length is 2,078 nucleotides, of which 38 represent the 5'-untranslated region, 54 identify a prepeptide of 18 amino acids, 18 identify a propeptide of 6 amino acids, 1,755 code for the known 585 amino acids of serum albumin, 189 make up the 3'-untranslated region and 24 are the poly(A) sequence. Nucleotides 5 to 15 (-34 to -24) in the 5'-untranslated region (Table 1) are complementary to a 3'-terminal region of eukaryotic 18S RNA [Azad, A.A. and Deacon, N.J. (1980) Nucl. Acids Res. 8, 4365-4376] and thus could represent a ribosome binding site:

(5')...T^CT C T T C T G T.....albumin mRNA
 (3')...G A G G A A G G C G U C C m₂⁶A m₂⁶A.....18S RNA

The translated portion of the mRNA sequence codes for the signal peptide and the main body of the albumin polypeptide chain. The

signal peptide is composed of a hydrophobic prepeptide of 18 amino acids and a basic propeptide of 6 amino acids (Table 1). Since prepeptides are removed from nascent secretory proteins (like albumin) in the endoplasmic reticulum, they are seen only in vitro in heterologous translation systems. As yet, they have not been found within cells [Judah, J.D. and Quinn, P.S. (1977) FEBS 11th Mtg., Copenhagen 50, 21-29; and Strauss, A.W., Donohue, A.M., Bennett, C.D., Rodkey, J.A. and Alberts, A.W. (1977) Proc. Natl. Acad. Sci. USA 74, 1358-1362]. This is the first report of the presence and the sequence of a prepeptide for human serum albumin. As it is with other secretory proteins, the conversion of proalbumin to albumin takes place in the Golgi vesicles, and the enzyme responsible for this cleavage is probably cathepsin B [Judah, J.D. and Quinn, P.S. (1978) Nature 271, 384-385]. This is also a first report on the sequence of the propeptide for normal human serum albumin.

At the 3'-end of the message, the putative polyadenylation signal sequence, AATAAA, is located 164 nucleotides downstream from the amino acid termination codon TAA and 16 nucleotides upstream from the beginning of the poly(A) sequence. Another characteristic sequence located near the polyadenylation site has been identified by Benoist, et al. [Benoist, C., O'Hare, K., Breathnach, R. and Chambon, P. (1980) Nucl. Acids Res. 8, 127-142]; the consensus sequence from several mRNAs was concluded as TTTTCACTGC. A similar sequence, TTTTCTCTGT, is located 19 nucleotides upstream from the AATAAA hexanucleotide in the human albumin mRNA (Table 1).

30

35

231 240 245 246 250 253 260 261
 val ser lys leu val thr asp leu thr lys val his thr glu cys cys his gly asp leu leu glu cys ala asp asp arg ala asp leu
 GTT TCC AAG TTA GTG ACA GAT CTT ACC AAA GTC CAC ACC GAA TGC TCC CAT GCA GAT CTG CTT GAA TGT GCT CAT CAC AGG GCG CAC CTT (890)

261 265 270 278 279 280 289 290
 ala lys tyr lle cys glu asn gln asp ser lle ser ser lys leu lys glu cys glu lys pro leu leu glu lys ser his cys lle
 CCC AAG TAT ATC TGT GAA AAT CAA GAT TCG ATC TCC AGT AAA CTG AAG GAA TGC TGT GAA AAA CCT CTG TTC GAA AAA TCC CAC TGC ATT (980)

291 300 303 306 310 316 320 321
 ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val cys lys asn tyr ala
 CCC CAA GTG CAA AAT GAT GAG ATG CCT GCT CAC TTG CCT TCA TTA GCT GCT CAT TTT GTT GAA AGT AAG CAT GTT TGC AAA AAC TAT GCT (1070)

321 330 333 336 340 340 350 351
 glu ala lys asp val phe leu gly met phe leu tyr glu tyr ala arg arg his pro asp tyr ser val val leu leu leu arg leu ala
 CAG GCA AAG CAT GTC TTC TTG GGC ATG TTT TTG TAT GAA TAT GCA AGA AGG CAT CCT GAT TAC TCT GTC GTC CTG CTG AGA CTT GCC (1160)

351 360 361 369 370 380 381
 lys thr tyr glu thr leu glu lys cys cys ala ala asp pro his glu cys tyr ala lys val phe asp glu phe lys pro leu
 AAG ACA TAT GAA ACC ACT CTA GAG MAG TCC TGT GCT GCT CCA GAT CCT CAT GAA TGC TAT GCC AAA GTG TTC GAT CAA TTT AAA CCT CCT (1250)

381 390 392 400 410 411
 val glu glu pro gln asn leu lle lys gln asn cys glu leu phe glu gln leu qly glu tyr lys phe gln asn ala leu leu val arg
 GTG GAA CAG CCT CAG AAT TTA ATC AAA CAA AAT TGT GAG CTT TTT CAG CAG CTT GCA GAG TAC AAA TTC CAG AAT GCG CTG TTA GTT CGT (1300)

411 420 423 426 430 437 438 440 441
 tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu qly lys val qly ser lys cys cys lys his
 TAC ACC AAG AAA GTA CCC CAA GTG TCA ACT CCA ACT CTT GTA CAG GTC TCA AGA AAC CTA GCA AAA GTG GGC AGC AAA TGT TGT AAA CAT (1430)

441 448 450 460 461 470 471
 pro glu ala lys arg met pro cys ala glu asp tyr leu ser val val leu asn gln leu cys val leu his glu lys thr pro val ser
 CCT GAA GCA AAA AGA ATG CCC TGT CCA GAA CAC TAT CTA TCC GTG GTC AAC CAG TTA TGT TGT GTC CAT GAG AAA ACC CCA GTA AGT (1520)

471 476 477 480 490 500 501
 asp arg val thr lys cys cys thr glu ser leu val asn arg arg pro cys phe ser ala leu glu val asp glu thr tyr val pro lys
 GAC AGA GTC ACC AAA TCC TGC ACA CAA TCC TTG GTG AAC AGG CCA CCA TCC TTT TCA GCT CTG GAA GTC GAT GAA ACA TAC GTT CCC AAA (1610)

501 510 514 520 530 531
 glu phe asn ala glu thr phe thr phe his ala asp lle cys thr leu ser glu lys glu arg gln lle lys lys gln thr ala leu val
 CAG TTT AAT GCT GAA ACA TTC ACC TTC CAT GCA GAT ATA TCC ACA CTT TCT GAG AAG GAG AGA CAA ATC AAG AAA CAA ACT GCA CTT GTT (1700)

35 531 540 550 558 559 560
glu leu val lys his lys pro lys ala thr lys glu gln leu lys ala val met asp phe ala ala phe val glu lys cys cys lys
GAG CTC GTG AAA CAC MAG CCC MAG GCA ACA AAA GAG CAA CTG AAA GCT GTT ATG CAT TTT CCT GCT TTT GTA GAG AAG TGC TGC AAG (1790)
561 567 570 580
ala asp asp lys glu thr cys phe ala glu glu gly lys lys leu val ala ala ser gln ala ala leu gly leu ter
GCT GAC GAT AAG GAG ACC TGC TTT GCC GAG GAG GGT AAA AAA CTT GTT GCT GCA AGT CAA GCT GCC TTA GGC TTA TAA CATCACATTTAAAG (1883)
ter ter
CATCTCAGCCTACCATGACAAATAAGCAGCAAAATGAAGATCAAAAGCTTATTCATCTGTTTTCTTTTCGTTGGTAAAGCCACACCCCTGCTAAAAAACATAAATTTCTTTAA (2M2)
TCATTTGCCTCTTTTCTGCTGCTTCAATTAATAAAAATGCAAGATCTAA..... 20AA (2078)

Following are examples which illustrate procedures, ~~including the best mode~~, for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

5 Example 1 Isolation of Messenger RNA

Human liver mRNA was obtained following the procedure of Chirgwin, et al [Chirgwin, J.M., Przybyla, A.E., MacDonald, R.J. and Rutter, W.J. (1979) Biochemistry 18, 5294-5299]. Immunoprecipitation of albumin containing polysomes was performed according to Taylor and
10 Tse [Taylor, J.M. and Tse, T.P.H. (1976) J. Biol. Chem. 251, 7461-7467]. In vitro translation of mRNA was carried out in a reticulocyte cell-free system, following the instruction of the manufacturer (New England Nuclear). The translation products were separated electrophoretically according to Laemmli [Laemmli, J.K. (1970) Nature 227,
15 680-685.

Example 2 Cloning Procedures

Double stranded cDNA was synthesized as described previously [Law, S., Tamaoki, T., Kreuzaler, F. and Dugaiczky, A. (1980) Gene 10, 53-61]. It was annealed to PstI-linearized pBR322 DNA [Bolivar, F.,
20 Rodriguez, R.L., Greene, P.J., Betlach, M.C., Heyneker, H.L., Boyer, H.W., Crossa, J.H. and Falkow, S. (1977) Gene 2, 95-113] that had been tailed with 15 dG residues/3'-terminus [Dugaiczky, A., Robberson, D.L. and Ullrich, A. (1980) Biochemistry 19, 5869-5873]. The annealed DNA was used to transform E. coli strain RR1, as detailed previously [Law, S., et al., ibid.]. The albumin clones were selected using the colony
25 hybridization method of Grunstein and Hogness [Grunstein, M. and Hogness, D.S. (1975) Proc. Natl. Acad. Sci. USA 72, 3961-3965], with [³²P]-labeled cDNA synthesized with the immunoprecipitated polysomal mRNA as template.

30 As shown in Example 5, plasmids pHA36 and pHA206 were deposited in E. coli HB101 hosts. The plasmids were obtained from E. coli RR1 hosts, described in this example, and transformed into E. coli HR101 by standard procedures well known to those of ordinary skill in this art. The E. coli RR1 hosts were lysed and then centrifuged to
35 separate the chromosomal DNA, cell DNA and plasmid DNA. The plasmid DNA, remaining in the supernatant, is precipitated with ethanol and the precipitate is resuspended in buffer, e.g., TCM (10mM Tris·HCl, pH 8.0, 10 mM CaCl₂, 10 mM MgCl₂). The cells for transformation are

prepared as follows: 120 ml of L-broth (1% tryptone, 0.5% yeast extract, 0.5% NaCl) are inoculated with an 18 hour culture of HR101 NRRL B-11371 and grown to an optical density of 0.6 at 600 nm. Cells are washed in cold 100 mM NaCl and resuspended for 15 minutes in 20 ml chilled 50 mM CaCl₂. Bacteria are then concentrated to one-tenth of this volume in CaCl₂ and mixed 2:1 (v:v) with annealed plasmid DNA, prepared as described above. After chilling the cell-DNA mixture for 15 minutes, it is heat shocked at 42°C for 2 minutes, then allowed to equilibrate at room temperature for ten minutes before addition of L-broth 10 times the volume of the cell-DNA suspension. Transformed cells are incubated in broth at 37°C for one hour before inoculating selective media (L-agar plus 10 µg/ml tetracycline) with 200 µl/plate. Plates are incubated at 37°C for 48 hours to allow the growth of transformants.

15 Example 3 Mapping of Restriction Endonuclease Sites

Restriction endonucleases were obtained from Bethesda Research Laboratories and New England Biolabs and were used according to the manufacturers' instructions. The digested DNA fragments were analyzed electrophoretically on agarose [Helling, R.B., Goodman, H.M. and Boyer, H.W. (1974) J. Virol. 14, 1235-1244] or acrylamide [Dingman, C., Fisher, M.P. and Kakefuda, T. (1972) Biochemistry 11, 1242-1250] gels.

20 Example 4 DNA Sequencing

DNA fragments were dephosphorylated with bacterial alkaline phosphatase (Worthington) and labeled at the 5'-ends with polynucleotide kinase (Boehringer-Mannheim) and γ [³²P]ATP. Following digestion with a second restriction endonuclease and electrophoretic separation of the fragments, DNA sequence determination was done according to the procedure of Maxam and Gilbert [Maxam, A. and Gilbert, W. (1980) Methods Enzym. 65, 499-560] and the degradation products were separated electrophoretically on 0.4 mm acrylamide gels as described by Sanger and Coulson [Sanger, F. and Coulson, R. (1978) FEBS Letters 87, 107-110].

30 Example 5 Recombinant Plasmids pHA36 and pHA206

As disclosed in Example 2, albumin clones were selected by hybridizing to the enriched albumin cDNA probe. Plasmid pHA36 contained the largest insert of an albumin cDNA sequence. Both plasmids pHA36 and pHA206 have been deposited in a viable E. coli host in the

permanent collection of the Northern Regional Research Laboratory (NRRL), U.S. Department of Agriculture, Peoria, Illinois, U.S.A. Their accession numbers in this repository are as follows:

HB101(pHA36) - NRRL B-12551

5 HB101(pHA206) - NRRL B-12550

E. coli HB101 is a known and widely available host microbe. Its NRRL accession number is NRRL B-11371.

NRRL B-12550 and NRRL B-12551 are available to the public. ~~upon the grant of a patent. It should be understood that the availability~~
10 of these deposits does not constitute a license to practice the subject invention in derogation of patent rights granted with the subject instrument by governmental action.

E. coli RR1 and E. coli HB101 are known and widely available host microbes. Their NRRL accession numbers are NRRL B-12186 and NRRL
15 B-11371, respectively.

pBR322 is a well known and widely available plasmid. It can be obtained from the following host deposit by standard procedures:

NRRL B-12014 - E. coli RR1 (pBR322).

YE6 is a well known and widely available yeast episomal plasmid.
20 It can be obtained from the following host deposit by standard procedures:

E. coli HB101 (YE6) - NRRL B-12093.

Example 6 Assembly of the Serum Albumin Gene

Assembling the pieces together is a straightforward task of re-
25 striction enzymology. There is only one MspI site in the overlapping DNA sequence of the two cDNA clones. Two enzymatic steps of (i) MspI digestion of the two DNAs, followed by (ii) the use of ligase, an enzyme that seals DNA fragments, will give the desired product. Although two other undesired DNA species will also be obtained in the
30 course of this recombination reaction, both of them will differ substantially in size. Thus, separation and isolation of the desired DNA species will be achieved.

The assembled DNA clone can be used to transform two types of cells:

- 35 (a) Escherichia coli
(b) Saccharomyces cerevisiae

(a) The vector of choice is plasmid pBR322, the same that has

been successfully used for cloning of the two fragmented pieces of the serum albumin cDNA.

(b) In order to transform yeast with the serum albumin structural gene sequence, the DNA must be inserted into one of the existing yeast plasmid vectors. This can be accomplished by taking advantage of the fact that several restriction endonuclease recognition sequences are absent from the cloned serum albumin DNA. Synthetic EcoRI DNA linkers can be ligated to the DNA fragment containing the serum albumin sequence followed by insertion (ligation) into one of the yeast plasmid vectors, e.g., YEp6, at the Eco RI cloning site. The fused chimeric plasmid can be used to transform yeast according to an established procedure [Hinnen, A., Hicks, J.B. and Fink, G.R. (1978) Proc. Natl. Acad. Sci. USA, 75, 1929]. YEp6 can be obtained from the NRRL repository, as disclosed supra.

15 Example 7 Expression of the Serum Albumin Gene

The main body of the structural gene will be transcribed by the E. coli or yeast enzymes. If little or no albumin is produced with the selected host, then an Escherichia coli promoter DNA sequence carrying an initiation codon, i.e., ATG, can be ligated at the beginning of the serum albumin structural gene. Such elements are known and available, e.g., lac promoter used for the expression of human interferon gene in E. coli [Proc. Natl. Acad. Sci. 77, 5230 (1980)]; source of promoter DNA [Proc. Natl. Acad. Sci. 76, 760 (1979)]. Also, see Nature, Vol. 281, October 18, 1979. It has already been documented that such Escherichia coli promoter sequences function well in the expression of foreign genes in Escherichia coli [Mercereau-Puijalon, O., Royal, A., Cami, B., Garapin, A., Krust, A., Gannon, I. and Kourilsky, P. (1978) Nature 275, 505; and Goeddel, D.V., Kleid, D.G., Bolivar, F., Heyneker, H.L., Yansura, D.G., Grea, R., Hirose, T., Kraszewski, A., Itakura, K., and Riggs, A. (1979) Natl. Acad. Sci. USA 76, 106]. For expression in yeast, see Rose, M., Casadaban, M.J. and Botstein, D. (1981) Proc. Natl. Acad. Sci. USA 78, 2460 and 4466.

25 Example 8 Screening of Clones Producing Albumin

Immunological methods can be used to detect small amounts of albumin made in a bacterium. Flat disks of flexible polyvinyl are coated with the IgG fraction from an immune serum and the disks are pressed onto an agar plate so that antigen released from an in situ lysed microbial colony can bind to the fixed antibody. The plastic

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disk is then incubated with the same total IgG fraction labeled with radioactive iodine so that other determinants on the bound antigen can in turn bind the iodinated antibody. Radioactive areas on the disk expose X-ray film during autoradiography and thus identify colonies producing the protein which is being screened for. Detailed protocols of this procedure have been published [Broome, S. and Gilbert, W. (1978) Proc. Natl. Acad. Sci. USA, 75, 2746]. The purification of human serum albumin can be accomplished by using procedures well known in the art. For example, procedures disclosed in a chapter by T. Peters: Purification and Properties of Serum Albumin, in: The Plasma Proteins, Putnam, Ed. Academic Press, New York, 1975, can be used.

The work described herein was all done in conformity with physical and biological containment requirements specified in the NIH Guidelines.

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CLAIMS

1. Plasmid pHA36, having a restriction endonuclease pattern as shown in the drawing.
- 5 2. Plasmid pHA206, having a restriction endonuclease pattern as shown in the drawing.
- 10 3. E. coli HB101 (pHA36) having the deposit accession number NRRL B-12551.
- 15 4. E. coli HB101 (pHA206) having the deposit accession number NRRL B-12550.
- 20 5. A microorganism modified to contain a nucleotide sequence coding for the amino acid sequence of human serum albumin; said nucleotide sequence is as follows:

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231 240 245 246 250 253 260
 val ser lys leu val thr asp leu thr lys val his thr glu cys his gly asp leu leu glu cys ala asp asp arg ala asp leu
 GTT TCC AAG TTA GTG ACA GAT CTT ACC AAA GTC CAC ACG GAA TGC TGC CAT GCA GAT CTG CTT GAA TGT GAT GAC AGG GCG GAC CTT (890)

261 265 270 278 279 280 289 290
 ala lys tyr ile cys glu asn gln asp ser ile ser ser lys leu lys glu cys oys qiu lys pro leu leu glu lys ser his cys lle
 GCC AAG TAT ATC TGT GAA AAT CAA GAT TCC ATC TCC AGT AAA CTG AAG GAA TCC TGT GAA AAA CCT CTG TTG GAA AAA TCC CAC TCC ATT (980)

291 300 306 310 316 320
 ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val oys lys asn tyr ala
 GCC GAA GTC GAA AAT GAT GAG ATG CCT GCT CAC TTG CCT TCA TTA CCT GCT CAT TTT GTT GAA AGT AAG GAT GTT TCC AAA AAC TAT GCT (1070)

321 330 336 340 350
 glu ala lys asp val phe leu glu met phe leu tyr glu tyr ala arg arg his pro asp tyr ser val val leu leu ero leu ala
 CAG GCA AAG GAT GTC TTC TTG GGC ATG TTT TTG TAT GAA TAT GCA AGA AGG CAT CCT GAT TAC TCT GTC CTG CTG ACA CTT GCC (1160)

351 360 361 369 370 380
 lys thr tyr glu thr leu glu lys oys ala ala asp pro his qiu cys tyr ala lys val phe asp qiu phe lys pro leu
 AAG ACA TAT GAA ACC ACT CTA GAG AAG TGC TGT GCC GCT CCA GAT CCT CAT GAA TGC TAT GCC AAA GTG TTC GAT GAA TTT AAA CCT CCT (1250)

381 390 392 400 410
 val glu glu pro gln asn leu ile lys gln asn cys glu leu phe qiu qln leu gly glu tyr lys phe qln asn ala leu leu val arg
 GTG GAA GAG CCT CAG AAT TTA ATC AAA CAA AAT TGT GAG CTT TTT GAG CAG CTT GCA GAG TAC AAA TTC CAG AAT CCG CTG TTA GTT CGT (1340)

411 420 430 437 438 440
 tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu qly lys val qly ser lys cys lys his
 TAC ACC AAG AAA GTA CCC CAA GTG TCA ACT CCA ACT CTT GTA GAG GTC TCA AGA AAC CTA GCA AAA GTG GCC ACC AAA TGT TGT AAA CAT (1430)

441 448 450 460 461 470
 pro glu ala lys arg met pro cys ala glu asp tyr leu ser val val leu asn gln leu cys val leu his qiu lys thr pro val ser
 CCT GAA GCA AAA AGA ATG CCC TGT GCA GAA GAC TAT CTA TCC GTG GTC AAC CAG TTA TGT GTG TTG CAT GAG AAA ACG CCA GTA AGT (1520)

471 476 477 480 490 500
 asp arg val thr lys cys thr glu ser leu val asn arg arg pro cys phe ser ala leu glu val asp qiu thr tyr val pro lys
 CAC ACA GTC ACC AAA TGC TGC ACA GAA TCC TTG GTG AAC AGG CCA CCA TCC TTT TCA GCT CTG GAA GTC GAT GAA ACA TAC GTT CCC AAA (1610)

501 510 514 520 530
 glu phe asn ala glu thr phe thr phe his ala asp ile cys thr leu ser qiu lys glu arg qln ile lys lys qln thr ala leu val
 GAG TTT AAT GCT GAA ACA TTC ACC TTC CAT GCA GAT ATA TCC ACA CTT TCT CAG AAG GAG ACA CAA ATC AAG AAA CAA ACT GCA CTT GTT (1700)

35 531 540 550 558 559 560
glu leu val lys his lys pro lys ala thr lys ala thr lys glu gln leu lys ala val met asp asp phe ala ala phe val glu lys eys eys lys
GAG CTC GTG AAA CAC AAG CCC AAG GCA ACA AAA GAG CAA CTG AAA CCT GTT ATG CAT TTT GCT GCT TTT GTA GAG AAG TGC AAG (1790)

561 567 570 580
ala asp asp lys glu thr cys phe ala glu glu qly lys lys leu val ala ala ser gln ala ala leu qly leu ter ter
CCT GAC GAT AAG CAG ACC TGC TTT GCC CAG CAG GGT AAA AAA CTT GTT GCT GCA AGT CAA GCT GCC TTA GCC TTA TAA CATCACATTTAAAG (1883)

ter ter
CATCTCACCCCTACCATCAGAAATAAGAGAAAGAAAATCAAGATCAAAAAGCCTTATTCATCTGTTTTCTTTTCGTTGCTAAAGCCACCCCTGCTAAAAAACATAAATTTCTTAA (2002)

TCATTTGCCICITTTTCTGCTGCTTCAATTAATAAAAAATGGAAAGAAATCTAA..... 20AA (2078)

231 val ser lys leu val thr asp leu thr lys val his thr glu cys cys his gly asp leu leu glu cys ala asp asp arg ala asp leu 260
 GTT TCC AAG TTA GTG ACA GAT CTT ACC AAA GTC CAC ACG GAA TGC TGC CAT GGA GAT CTG CTT GAA TGT CCT GAT CAC ACG CCG CAC CTT (890)
 240 245 246 250 253
 261 ala lys tyr lle cys glu asn gln asp ser lle ser ser lys leu lys glu cys qiu lys pro leu leu qiu lys ser his cys lle 289 290
 CCC AAG TAT ATC TGT GAA AAT CAA GAT TCG ATC TCC AGT AAA CTG AAG GAA TGC TGT GAA AAA CCT CTG TTG GAA AAA TCT CAC TGC ATT (980)
 270 278 279 280
 291 ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val cys lys asn tyr ala 320
 GCC GAA GTG GAA AAT GAT GAG ATG CCT CCT GAC TTG CCT TCA TTA CCT CCT GAT TTT GTT GAA AGT AAG GAT GTT TGC AAA AAC TAT CCT (1070)
 300 316
 321 glu ala lys asp val phe leu gly met phe leu tyr glu tyr ala arg arg his pro asp tyr ser val val leu leu leu arg leu ala 350
 GAG GCA AAG GAT GTC TTC TTG GGC ATG TTT TTG TAT GAA TAT GCA AGA ACG CAT CCT GAT TAC TCT GTC GTG CTG CTG ACA CTT GCC (1160)
 330 340
 351 lys thr tyr glu thr thr leu glu lys cys cys ala ala asp pro his qiu cys tyr ala lys val phe asp glu phe lys pro leu 380
 AAG ACA TAT GAA ACC ACT CTA GAG AAG TGC TGT GCC GCT GCA GAT CCT CAT GAA TGC TAT GCC AAA GTG TTC GAT GAA TTT AAA CCT CCT (1250)
 360 361 369 370
 381 val glu glu pro gln asn leu lle lys gln asn cys glu leu phe qiu qin leu qiu gly glu tyr lys phe qin asn ala leu leu val arg 400 410
 GTG GAA GAG CCT CAG AAT TTA ATC AAA CAA AAT TGT GAG CTT TTT GAG CAG CTT GCA GAG TAC AAA TTC CAG AAT GCC CTG TTA GTT CGT (1380)
 390 392 400
 411 tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu qly lys val qly ser lys cys cys lys his 430 437 438 440
 TAC ACC AAG AAA GTA CCC CAA GTG TCA ACT CCA ACT CTT GTA GAG GTC TCA ACA AAC CTA GCA AAA GTG GCC ACC AAA TGT TGT AAA CAT (1830)
 420 430
 441 pro glu ala lys arg met pro cys ala glu asp tyr leu ser val val leu asn gln leu cys val leu his qiu lys thr pro val ser 460 461 470
 CCT GAA GCA AAA ACA ATG CCC TGT GCA GAA GAC TAT CTA TCC GTG GTC CTG AAC CAG TTA TGT GTG TTG CAT GAG AAA ACC CCA GTA AGT (1520)
 448 450 460 461
 471 asp arg val thr lys cys cys thr glu ser leu val asn arg pro cys phe ser ala leu glu val asp qiu thr tyr val pro lys 490 500
 GAC ACA GTC ACC AAA TCC TCC ACA GAA TCC TTG GTG AAC AGG CGA CCA TGC TTT TCA GCT CTG GAA GTC GAT GAA ACA TAC GTT CCC AAA (1610)
 476 477 480 490
 501 glu phe asn ala glu thr phe thr phe his ala asp lle cys thr leu ser qiu lys glu arg qin lle lys lys qin thr ala leu val 510 514 520 530
 CAG TTT AAT GCT GAA ACA TTC ACC TTC CAT GCA GAT ATA TCC ACA CTT TCT CAG AAG GAG ACA CAA ATC AAG AAA CAA ACT GCA CTT GTT (1700)

7. Nucleotide sequence coding for the prepeptide of human serum albumin, said nucleotide sequence is as follows:

5
10
15
20
25
30
35

p r o

-10

Met lys trp val tlu phe lle ser leu leu phe leu phe ser
ATG AAG TGG GTA ACC TTT ATT TCC CTT CTT TTT CTC TTT AGC (30)

p r o

-1

-6

ser ala tyr ser arg gly val phe arg arg
TCC CCT TAT TCC ACC GGT GTC TTT CGT CCA

GCCTTCTCTCTGTCACCCACAGCCCTTTGGCACA

231 240 245 246 250 253 260 265
 val ser lys leu val thr asp leu thr lys val his thr glu cys cys his gly asp leu leu glu cys ala asp asp arg ala asp leu
 GTT TCC AAG TTA GTG ACA GAT CTT ACC AAA GTC CAC ACC GAA TGC TGC CAT GGA GAT CTG CTT GAA TGT CCT CAT CAC AGG CCG GAC CTT (890)

261 265 270 278 279 280 289 290
 ala lys tyr ile cys glu asn gln asp ser ile ser ser lys leu lys glu cys glu lys pro leu leu glu lys ser his cys ile
 GCC AAG TAT ATC TGT GAA AAT CAA GAT TCG ATC TCC AGT AAA CTG AAG GAA TCC TGT GAA AAA CCT CTG TTG GAA AAA TCC CAC TGC ATT (980)

291 300 310 316 320
 ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val cys lys asn tyr ala
 GCC GAA GTG GAA AAT CAT CAG ATG CCT GCT CAC TTG CCT TCA TTA CCT GCT CAT TTT GTT GAA AGT AAG CAT GTT TGC AAA AAC TAT CTT (1070)

321 330 340 350
 glu ala lys asp val phe leu gly met phe leu tyr glu tyr ala arg arg his pro asp tyr ser val val leu leu leu arg leu ala
 GAG GCA AAG GAT GTC TTC TTG GCC ATG TTT TTG TAT GAA TAT GCA AGA AGG CAT CCT GAT TAC TCT GTC GTG CTG ACA CTT GCC (1160)

351 360 361 369 370 380
 lys thr tyr glu thr thr leu glu lys cys cys ala ala asp pro his glu cys tyr ala lys val phe asp glu phe lys pro leu
 AAG ACA TAT GAA ACC ACT CTA CAG AAG TCC TGT GCC CCT CCA CAT CCT CAT GAA TGC TAT GCC AAA GTG TTC CAT GAA TTT AAA CCT CCT (1250)

381 390 392 400 410
 val glu glu pro gln asn leu ile lys gln asn cys glu leu phe glu gln leu qly glu tyr lys phe gln asn ala leu leu val arg
 GTG GAA GAG CCT CAG AAT TTA ATC AAA CAA AAT TGT CAG CTT TTT CAG CAG CTT GCA GAG TAC AAA TTC CAG AAT CCG CTG TTA GTT CGT (1340)

411 420 430 437 438 440
 tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu qly lys val qly ser lys cys lys his
 TAC ACC AAG AAA GTA CCC CAA GTG TCA ACT CCA ACT CTT GTA CAG GTC TCA ACA AAC CTA GCA AAA GTG GCC AGC AAA TGT TGT AAA CAT (1430)

441 448 450 460 461 470
 pro glu ala lys arg met pro cys ala glu asp tyr leu ser val val leu asn gln leu cys val leu his glu lys thr pro val ser
 CCT GAA GCA AAA ACA ATG CCC TGT GCA GAA CAC TAT CTA TCC GTG GTC CTG AAC CAG TTA TGT GTG TTG CAT CAG AAA ACC CCA GTA AGT (1520)

471 476 477 480 490 500
 asp arg val thr lys cys thr glu ser leu val asn arg pro cys phe ser ala leu glu val asp glu thr tyr val pro lys
 GAC AGA GTC ACC AAA TCC TCC ACA GAA TCC TTG GTG AAC AGG CGA CCA TCC TTT TCA CCT CTG GAA GTC GAT GAA ACA TAC CTT CCC AAA (1610)

501 510 514 520 530
 glu phe asn ala glu thr phe thr phe his ala asp ile cys thr leu ser glu lys glu arg gln ile lys lys gln thr ala leu val
 GAG TTT AAT CCT GCT GAA ACA TCC ACC TTC CAT GCA GAT ATA TCC ACA CTT TCT CAG AAG GAG AGA CAA ATC AAG AAA CAA ACT GCA CTT GTT (1700)

231	240	243 246	250	253	260
val ser lys leu val thr asp leu thr lys val his thr glu cys oys his gly asp leu leu glu cys ala asp asp arg ala asp leu	240	243 246	250	253	260
GTT TCC AAG TTA GTG ACA GAT CTT ACC AAA GTC CAC ACG GAA TGC TCC CAT GGA GAT CTG CTT GAA TGT GCT GAT GAC AGG GCG GAC CTT (890)	240	243 246	250	253	260
261	270	278 279 280	289 290		
ala lys tyr ile cys glu asn gln asp ser ile ser ser lys leu lys glu cys qiu lys pro leu leu glu lys ser his cys ile	270	278 279 280	289 290		
CCC AAG TAT ATC TGT GAA AAT CAA GAT TCG ATC TCC AGT AAA CTG AAG GAA TCC TGT GAA AAA CCT CTG TTC GAA AAA TCT CAC TGC ATT (980)	270	278 279 280	289 290		
291	300	310	316	320	
ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val cys lys asn tyr ala	300	310	316	320	
GCC GAA GTG GAA AAT GAT GAG ATG CCT GCT GAC TTG CCT TCA TTA GCT GCT GAT TTT GTT GAA AGT AAG GAT GTT TGC AAA AAC TAT GCT (1070)	300	310	316	320	
321	330	340	350		
glu ala lys asp val phe leu gly met phe leu tyr glu tyr ala arg arg his pro asp tyr ser val val leu leu leu arg leu ala	330	340	350		
GAG GCA AAG GAT GTC TTC TTG GCC ATG TTT TTG TAT GAA TAT GCA AGA ACG CAT CCT GAT TAC TCT GTC GTG CTG CTG AGA CTT GCC (1160)	330	340	350		
351	360 361	369 370	380		
lys thr tyr glu thr thr leu glu lys oys cys ala ala asp pro his qiu cys tyr ala lys val phe asp alu phe lys pro leu	360 361	369 370	380		
AAG ACA TAT GAA ACC ACT CTA GAG AAG TCC TGT GCC GCT GCA GAT CCT CAT GAA TGC TAT GCC AAA GTG TTC GAT GAA TTT AAA CCT CCT (1250)	360 361	369 370	380		
381	390	392	400	410	
val glu glu pro gln asn leu ile lys gln asn cys glu leu phe qiu leu qiu glu tyr lys phe gln asn ala leu leu val arg	390	392	400	410	
GTC GAA GAG CCT CAG AAT TTA ATC AAA CAA AAT TGT GAG CTT TTT GAG CAG CTT GCA GAG TAC AAA TTC CAG AAT CCG CTG TTA GTT CGT (1360)	390	392	400	410	
411	420	430	437 438	440	
tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu qiy lys val qiy ser lys cys oys lys his	420	430	437 438	440	
TAC ACC AAG AAA GTA CCC CAA GTG TCA ACT CCA ACT CTT GTA GAG GTC TCA ACA AAC CTA GCA AAA GTG GCC AGC AAA TGT TGT AAA CAT (1630)	420	430	437 438	440	
441	448	450	460 461	470	
pro glu ala lys arg met pro cys ala glu asp tyr leu ser val val leu asn gln leu cys val leu his qiu lys thr pro val ser	448	450	460 461	470	
CCT GAA GCA AAA ACA ATG CCC TGT GCA GAA GAC TAT CTA TCC GTG GTC CTG AAC CAG TTA TGT GTG TTC CAT GAG AAA ACG CCA GTA AGT (1520)	448	450	460 461	470	
471	476 477	480	490	500	
asp arg val thr lys oys cys thr glu ser leu val asn arg arg pro oys phe ser ala leu glu val asp qiu thr tyr val pro lys	476 477	480	490	500	
GAC AGA GTC ACC AAA TCC TCC ACA TCC TCG ACA GAA TCC TTG GTG AAC ACG CCA TCC TTT TCA GCT CTG GAA GTC GAT GAA ACA TAC GTT CCC AAA (1610)	476 477	480	490	500	
501	510	514	520	530	
glu phe asn ala glu thr phe thr phe his ala asp ile cys thr leu ser glu lys glu arg qin ile lys lys qin thr ala leu val	510	514	520	530	
GAG TTT AAT GCT GAA ACA TTC ACC TTC CAT CCA GAT ATA TCC ACA CTT TCT GAG AAG GAG ACA CAA ATC AAG AAA CAA ACT GCA CTT GTT (1700)	510	514	520	530	

10. A nucleotide sequence according to any of claims 6 to 9, in essentially pure form.
11. A DNA transfer vector comprising a nucleotide sequence as defined in claim 5.
- 5 12. A DNA transfer vector according to claim 11, transferred to and replicated in a micro-organism.
13. A DNA transfer vector according to claim 12, which is a plasmid.
14. A DNA transfer vector according to claim 13,
10 wherein the plasmid is pBR322 or YEp6.
15. A process for preparing human serum albumin, which comprises culturing a micro-organism according to claim 5.
16. A DNA transfer vector according to any of
15 claims 12 to 14, or a process according to claim 15, wherein the micro-organism is a bacterium or yeast.
17. A vector or process according to claim 16, wherein the bacterium or yeast is E. coli or Saccharomyces cerevisiae.

